

21 **Abstract:** The search of new solvents is currently focused on Deep Eutectic Solvents (DES). However,
22 there are not many ecotoxicological studies in different biomodels of DES that allow knowing how these
23 chemicals affect to the environment along the trophic chain. In this manuscript, two DES at different
24 proportion of water have been prepared and characterized from the ecotoxicological point of view. These
25 solvents are: glucose:chloride choline (2:5) and sorbitol:chloride choline (3:2) at different content of water.
26 To carry out the ecotoxicological study, three biomodels have been used: bacteria *Allivibrio fisheri* (*A.*
27 *fisheri*), crustacean *Daphnia magna* (*D. magna*) and algae *Raphidocelis subcapitata* (*R. subcapitata*). The
28 obtained results show that the ecotoxicity of these chemicals depends on the biomodel used and the amount
29 of water, being toxicity values lower for chemicals with higher water content. However, it is important to
30 highlight that the ecotoxicity for all chemicals is quite low with effective concentrations, EC₅₀ values above
31 1000 mg/L in all the studied cases.

32

33 **Keywords:** ecotoxicity, DES, glucose sorbitol, *Allivibrio fisheri*, *Daphnia magna*, *Raphidocelis*
34 *subcapitata*.

35

36 1. Introduction

37 Deep Eutectic Solvents (DES) and Natural Deep Eutectic Solvents (NADES) are mixtures formed
38 by two or more chemicals at solid state at room temperature but, when combined in a specific molar ratio,
39 form a liquid solution. This is mainly due the strong and complex network of intermolecular forces
40 established that cause a sharp diminution of the melting point (Laura Lomba 2021a).

41 DES can be considered as “design solvents” since the possibility of combinations of different
42 components and compositions is almost unlimited. The goal is setting the right mixture for obtaining the
43 physicochemical properties required for a specific chemical processes (Mitar et al. 2019).

44 DES are formed by a hydrogen bond acceptor (HBA) and a hydrogen bond donor (HBD) and the
45 typical components are natural chemicals such as: organic acids, alcohols, sugars, amino acids, urea, small
46 hydrophilic molecules or natural metabolites among others (Nystedt et al. 2021).

47 DES are usually low cost, are also easy to prepare (Benvenuti et al. 2020, Benvenuti et al. 2019),
48 present low volatility, are biodegradable (Wils et al. 2021) and normally non-reactive with water. The label
49 of non-toxic mixtures has to be reviewed and lately there is some controversy in the scientific community
50 in this regard; there are an important number of studies that show low toxicity of eutectic mixtures for the
51 environment or human both *in vitro* and *in vivo* studies (Laura Lomba 2021b), while some others studies
52 demonstrate the toxicity of DES above a certain limit of concentration (Torregrosa-Crespo et al. 2020).
53 Nevertheless, the toxicity of DES depends on several factors that include not only the nature of the
54 components of the mixture but also the organisms on which toxicity is studied, their pre-adaptation periods
55 or even the sterilization processes carried out during the measurement of toxicity. This may be caused by
56 the acidification of the medium caused by the DES hydrolysis (Torregrosa-Crespo et al. 2020). Thus, it is
57 required more specific studies about the effect of DES on different endpoints, covering several trophic
58 levels, organisms, cells or tissues and the use of more appropriate toxicological techniques (Marchel et al.
59 2022b, Ruesgas-Ramon et al. 2017).

60 It is also said that present good physicochemical properties for several chemical processes thanks
61 to their wide polarity range and low vapour pressure or chemical stability, among others (Benvenuti et al.
62 2020, Laura Lomba 2021a, Laura Lomba 2021b, Silva et al. 2019). For these reasons DES can be used as
63 stationary phases in chromatography (Momotko et al. 2021, 2022) and membranes (Castro-Munoz et al.
64 2022, Khajavian et al. 2022) and additionally, they are attractive for several industrial processes such as
65 biocatalysis (Zhou et al. 2021), cryopreservation (Craveiro et al. 2021), biochemical applications (Silva et
66 al. 2019, Yang 2019), extraction medium (Ruesgas-Ramon et al. 2017, Rukavina et al. 2021), biological
67 assays (Liu et al. 2018), pharmaceutical industry (Liu et al. 2021, Morrison et al. 2009) or cosmetic
68 industry (Benoit et al. 2021).

69 However, the main barrier to the widespread use of DES is the lack of information regarding
70 various physicochemical and toxic properties. This limitation arises from the very nature of the DES; the
71 multiple combinations between components and compositions (AlOmar et al. 2016, Lapena et al. 2020,
72 Lapena et al. 2019b, a) makes difficult to have a complete set of properties that allow knowing the specific
73 characteristics of a mixture (components and concentration) (Li et al. 2022) or even to predict the behaviour.

74 However, it is important notice that the thermal stability of DES can be diminished at high
75 temperature. Most of the studies about this issue analyse the thermal stability of DES using the onset

76 decomposition temperatures (Tonset). These temperatures, that were normally obtained from dynamic
77 thermogravimetry (TGA) under different experimental conditions that, in general, lead to the
78 overestimation of the onset decomposition temperature. It is important to bear in mind that there are some
79 scientific studies related to the thermal stability of DES that have found, in some cases, that DES can
80 decompose into HBA and HBD due to the weakening of the hydrogen bonds. This process can occur in two
81 steps: the first one, the DES component with a lower boiling point or little stability undergoes volatilization
82 or decomposition (normally HBD) and the second one, the decomposition of the other DES at a higher
83 temperature (normally the HBA). Thus, the role of the hydrogen bonds in the thermal stability seems to be
84 determinant. (Gutierrez et al. 2010, Marchel et al. 2022a). Additionally, it is important to note that the
85 toxicity of DES may vary when the hydrogen bond net is modified; new supramolecular structures can be
86 created, and thus, the toxicological activity may be also different.(Gutierrez et al. 2010, Marchel et al.
87 2022b, a)

88 With the aim of increasing the information available on these substances, we have previously
89 explored the ecotoxicity of some of them: reline, ethaline and glyceline towards several aquatic biomodels
90 (algae, crustaceans and bacteria)(Lapena et al. 2021). We founded that the ecotoxic effect clearly depended
91 on the studied biomodel and the type of test used as a tool to monitor toxicity. Furthermore, the water
92 content did not follow the expected trend and studied DES behaved erratically with regards to the water
93 composition(Lapena et al. 2021).

94 Thus, with the objective of deepening the knowledge of the ecotoxic behaviour of DES, in this
95 work we have studied the ecotoxicity of several eutectic mixtures containing sugars and chloride choline
96 at different concentrations of water: glucose:chloride choline (2:5) and sorbitol:chloride choline (3:2) with
97 different water content. Concretely, we have obtained the toxic effect that these mixtures produce in various
98 aquatic organisms, covering the trophic chain: bacteria *Allivibrio fisheri* (*A. fisheri*), crustacean *Daphnia*
99 *magna* (*D. magna*) and algae *Raphidocelis subcapitata* (*R. subcapitata*).

100 Besides, it has been previously reported that viscosity is an important properties key to explain
101 toxicity in some cells. An increase of toxicity is normally associated with higher cell lethality rates(Hayyan
102 et al. 2016). For that reason, we have also measured the kinematic viscosity of the studied DES. Toxicity
103 results have first been analysed in an independent way, considering both the DES and the biomodel. They
104 were then considered together and the final conclusions about the ecotoxic behavior have been reached.

105

106 **2. Material and methods**

107 *2.1 Chemicals and preparation of Deep Eutectic Solvents*

108 In Table 1, the used chemicals for the DES preparation are gathered. Choline chloride has been
109 dried under vacuum for 24 h prior to use. Mixtures with water have been prepared considering the previous
110 amount of water of each of the components of the mixtures with Milli-Q water (resistivity less than 18.2
111 M Ω ·cm) using a Sartorius Entris 5201-1S balance (uncertainty $\pm 10^{-1}$ g). Once each substance has been
112 weighed, has been transferred to a closed flask (250 ml) with a magnetic nucleus inside. The flask has been
113 placed on a heating plate and heated to a temperature of approximately 80°C with continuous stirring. The
114 process has been completed within one hour and a homogeneous liquid has been observed. In Table 2,
115 studies DES, composition, and abbreviation are shown.

116 Table 1. Characteristics of the used chemicals

Name	CAS	Supplier	Mass fraction purity	Formula	Molar mass (g/mol)
Glucose anhydrous	50-99-77	Panreac	0.975	C ₆ H ₁₂ O ₆	180.16
Sorbitol	50-70-4	Sigma-Adrich	0.980	C ₆ H ₁₄ O ₆	182.17
Choline Chloride	67-48-1	Sigma-Aldrich	0.993	C ₅ H ₁₄ ClNO	139.63

117

118 Table 2. Studied DES: composition and abbreviation

HBD	HBA	Add-on	Molar ratio	Abbreviation
			5:2:5	Glu5
Glucose	Choline chloride	Water	5:2:7.5	Glu7.5
			5:2:10	Glu10
			2:3:5	Sor5
Sorbitol	Choline chloride	water	2:3:7.5	Sor7.5
			2:3:10	Sor10

119

120 2.2 Viscosity measurements

121 Kinematic viscosities, ν , have been obtained using a Schoot-GeräteAVS-440 automatic measuring
 122 unit along with several Ubbelohde capillary viscosimeters. The uncertainty of the time flow measurements
 123 is 0.01 s, and kinetic energy corrections have been applied to the experimental data. The temperature has
 124 been controlled at 25°C by means of a Schoot-Geräte CT 1150/2 thermostat, with a temperature uncertainty
 125 of 0.01K.

126

127 2.3 A. *fischeri* ecotoxicity test

128 The lyophilized *A. fischeri* (strain NRRL-B-11177 and reference 945006) have been supplied by
 129 Macherey-Nagel. This test has been based on UNE-11348-3(UNE-11348-3 2009). All solutions have been
 130 prepared with 2% of NaCl and the pH has been adjusted using 0.1 M HCl or 0.1 M NaOH in 2 % NaCl.
 131 Additionally, positive controls such as phenol and zinc sulfate have been used(Jennings et al. 2001) and
 132 negative control has been culturing medium (Biofix Lumi medium for freeze-dried luminous bacteria by
 133 DIN EN ISO 11348-3, Macherey-Nagel, Duren, Germany).

134 Luminescence has been measured in acute mode (Biotox B) using a Biofix Lumi-10 luminometer
 135 (MachereyNagel) after 30 minutes of exposure and at 15°C. Three replicates for each control have been
 136 tested. More details of this test can be found in our previous works(Garcia et al. 2015, Lomba et al. 2014).

137

138 2.4 D. *magna* ecotoxicity test

139 *D. magna* ecotoxicity test have been based on the guidelines OECD 202(202 2004). The *D. magna*
 140 ehippia (Toxkit, Daphtokit F Magna, ref. DM090812) have been stored at 4°C and purchased from
 141 Vidrafoc. The eggs have been incubated with culturing medium considering the specifications of the
 142 supplier, during 72 h at 22°C with 6000 lux in a Toxkit CH-0120D-AC/DC incubator (supplied by Ecotest)

143 and fed with *Spirulina* algae 2 h before starting the experiment. The pH has been adjusted 7-7.5 before
144 exposure. Furthermore, positive control (sodium dichromate) and negative controls have been tested.

145 A total of 20 newborn daphnids (aged less than 24 h) have been exposed to the different dilutions
146 of the DES in dark for 24 h at 20°C per compound and concentration. The crustaceans have been separated
147 into four groups of five organisms, four replicates per concentration exposure. The test has been repeated
148 in triplicate. Daphnids have been considered immobilized if they have been not able to swim for 15 s after
149 agitation. More details of this protocol can be found in bibliography(Perales et al. 2017).

150

151 2.5 *R. subcapitata* ecotoxicity test

152 *R. subcapitata* test have been carried out according to OECD 201(OECD 1984). Algae have been
153 purchased from ECOTEST (SC2B1214). All the experiments have been repeated in triplicate. Moreover,
154 culturing medium and de-mobilization of algae have been prepared using the supplier specifications.

155 The algae cells have been incubated at 23°C in 100 mL beaker containing 50 mL of culturing
156 medium with 10000 lux of illumination. Cells densities of have been correlated with optical density (OD)
157 at 670 nm basing on the supplier specifications. The algae initial concentration has been $3 \cdot 10^5$ cells/ml.

158 The pH of the solutions has been adjusted in the range of 7.9 and 8.2 using 0.1 M NaOH or 0.1 M
159 HCl solutions. The assay has been carried out in a 96 well plate. To minimize evaporation, the outer wells
160 of the plate have been filled with 0.3 mL of distilled water and only the inner wells have been filled with
161 controls and test solutions. The initial OD has been measured at 670 nm. The well plate has been incubated
162 at 23°C for 72 hours. Next, all plates have been re-suspended to avoid the settling of algae and the final OD
163 has been measured.

164

165 2.6 Statistics treatment

166 Data have been analysed using a non-linear regression-representing log (inhibitor) versus
167 inhibition.

168 To obtain the EC₅₀ values of each compound, the experimental results have been fitted to the
169 following function:

$$170 \quad \% I = \frac{100}{(1 + 10^{(a - \log C)^b})} \quad (4)$$

171 where %I denotes % inhibition of luminescence for *A. fischeri*, % of immobilization for *D. magna* and %
172 of inhibition of growth for *R. subcapitata*; C is the concentration in mg/L and Log EC₅₀ and *a* are the
173 adjustable parameters.

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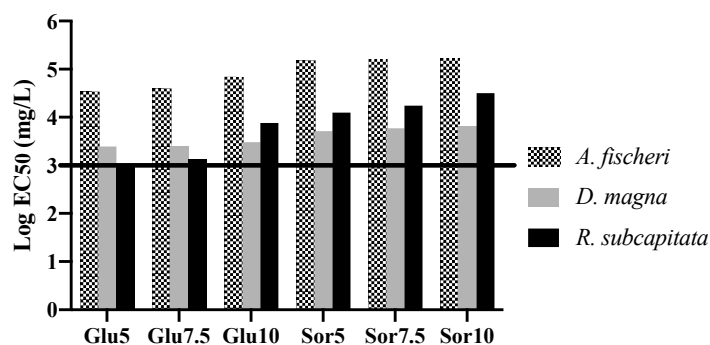
175 3. Results and discussion

176 In this section, the obtained results for the studies compounds are shown. Regarding to the
177 prepared DES, it is important to note that all DES formed by sorbitol (Sor5, Sor7.5 and Sor10) and glucose
178 (Glu10) have been obtained in a unique a clear phase at room temperature (25°C), however, in the case of
179 glucose DES (Glu5, Glu7) it has been necessary to slightly heat (30°C) to obtain that liquid phase before
180 the tests.

181 In this section, results have been analyzed from the perspective of the DES (or DES + water
182 mixtures) itself, as a whole. However, it is important to revise the nature of the components of the eutectic
183 mixtures from the toxicological point of view. For instance, choline chloride is a known component of
184 Vitamin B, and plays important metabolic functions; choline is usually the chosen cellular raw material for
185 the synthesis of cellular phospholipidic membranes (phosphatidylcholine and sphingomyelin) (Hayyan et
186 al. 2016). On the other hand, from a cellular perspective glucose and sorbitol are essential carbohydrates;
187 their metabolism provides the required energy for different cellular functions. After adsorption, glucose
188 undergo glycolysis when energy is required in the cell, otherwise is stored as glycogen. The sorbitol
189 metabolism is an accessory pathway in the glucose route. Through a chain of reactions, the sorbitol is an
190 intermediate that converts glucose into fructose. The glycolytic pathway for glucose and fructose leads to
191 synthesis of nucleic acids through the pentose phosphate pathway, the energy production in the
192 mitochondria through tricarboxylic acid pathway and the fatty acid synthesis through the lipogenesis
193 process. For these reasons, it is expected high cellular tolerance of the studied DES formed by choline and
194 these sugars. However, during the eutectic mixture formation process and the establishment of the hydrogen
195 bond network, an important modification of the chemical and physical properties is made, and this includes
196 the toxicological behavior of the eutectic mixtures.

197 Returning to the analysis of the results obtained by the DES studied in the biomodels, values of
198 EC_{50} towards *A. fischeri*, *D. magna* and *R. subcapitata* are shown in Table 3. For all the studied chemicals,
199 there is a dependence relationship between the toxicity and the concentration; *i.e.*, the toxic effect increases
200 as the concentration does. The general toxicity trend observed for all biomodels is the following: Glu 5 >
201 Glu 7.5 > Glu10 > Sor5 > Sor 7.5 > Sor10. In Figure 1, the toxicity of obtained values for all the studied
202 biomodels are presented. The results indicate that the most sensitive species was *D. magna* followed by *R.*
203 *subcapitata* and *A. fischeri*.

204 It is important to highlight that none of the studies mixtures can be considered toxic to the
205 environment (values of EC_{50} do not exceed the general limit of 1000 mg/L). The Globally Harmonized
206 System of classification And labelling of Chemicals (GHS) (Nations 2021) has been used to categorize the
207 toxicity for substances hazardous to the aquatic environment, Short-term (acute) as follow: Acute 1 (EC_{50}
208 ≤ 0.1 mg/L), acute 2 ($EC_{50} > 1$ but ≤ 10 mg/L), acute 3 ($EC_{50} > 10$ but ≤ 100 mg/L). For this GHS
209 classification, algae, crustacea and fish, are considered surrogate species. However, data on other organisms
210 may also be considered and the classification can be applied too. The ecotoxicity results obtained in this
211 work indicates that for all studied mixtures there is no need to classify for short-term (acute) hazard. It is
212 worth mentioning the case of *A. fischeri* where some mixtures exhibited values of EC_{50} higher than 100000
213 mg/L indicating very low toxicity to this biomodel (Figure 1).



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Figure 1. Comparison of the obtained results for each biomodel. The solid line at $\text{Log EC}_{50} = 3$ mg/L correspond to the general toxicity limit 1000 mg/L.

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The lipophilia is one of the key properties traditionally used to explain the toxic behavior: the higher lipophilic character, the higher ability of the molecules to cross through the biological membranes and produce a whatever effect on the activity. In this case, focusing of the sugars forming part of the DES, glucose and sorbitol, our toxicity results are in close agreement with lipophilic trend described by their corresponding octanol-water partition coefficients, $\log K_{ow} = -3,17$ and -4.67 respectively (Chemspider, accessed at April 13rd 2022).

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In Table 3, values of kinematic viscosity are also gathered. For these mixtures, the viscosity has been higher than the viscosity of some typical molecular liquids such as water or some other low molecular weight organic solvents. In the case of glucose mixtures, the higher viscosity has been obtained for Glu5 followed by Glu7 and Glu10. In the case of DES containing sorbitol, the trend is the same as for glucose mixtures, being the most viscous Sor5 followed by Sor7.5 and Sor10. This property is clearly affected by the nature of its components, temperature, or molar ratio. The content of water is important too, not only in physicochemical properties but also in the integrity of DES changing their supramolecular interactions (El Achkar et al. 2021). As mentioned before, viscosity and content of water may explain the obtained results; it has been observed that high viscosity are associated with increased lethality (Hayyan et al. 2016).

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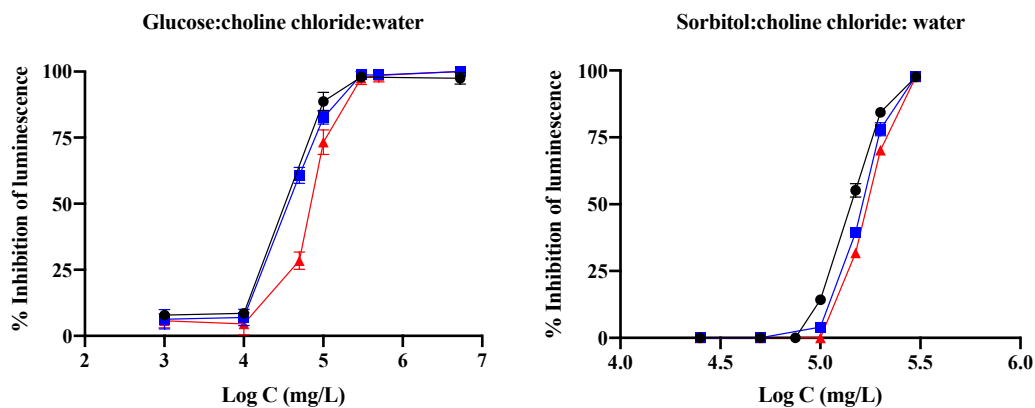
Table 3. Values of EC_{50} (mg/L) for *A. fischeri*, *D. magna* and *R. subcapitata* and values of kinematic viscosity at 25°C.

DES	<i>A. fischeri</i>	<i>D. magna</i>	<i>R. subcapitata</i>	Kinematic Viscosities
	30 min	24 h	72 h	ν , (mm/s)
Glu5	34196 ± 3965	2433 ± 31	1021 ± 122	787.69
Glu7.5	39955 ± 2455	2527 ± 22	1305 ± 114	375.22
Glu10	69591 ± 3073	3048 ± 24	7544 ± 475	185.02
Sor5	149748 ± 1279	5153 ± 85	12496 ± 663	3333.9
Sor7.5	161857 ± 812	5942 ± 283	17545 ± 295	613.01
Sor10	171209 ± 789	6557 ± 242	31325 ± 131	503.51

236

237 *A. fischeri* bacteria is Gram-negative, flagellated, non-pathogenic and rod-shaped bacterium,
 238 which is distributed in marine environments (Abbas et al. 2018). The bioluminescence mechanism is based
 239 in two substrates: luciferin and a long chain fatty aldehyde. Exogenous agents (reduced) produce the
 240 reduction of FMN (flavin mononucleotide oxidated) to FMNH₂ (flavin mononucleotide reduced) through
 241 luciferase (flavin mono-oxygenase oxidoreductase) action which reacts with oxygen forming an
 242 intermediate chemical called 4a-peroxy-flavin. This compound oxidizes the fatty aldehyde forming its
 243 corresponding acid and a luciferase–hydroxyflavin complex. This intermediate is decomposed slowly and
 244 emitting a blue-green light with its highest intensity at 490 nm (Meighen 1991). This process is completely
 245 linked to respiration, through the electron transport chain, and gives an idea about the metabolic status as a
 246 chemical toxicity. The presence of toxic substances diminishes the resultant luminescence. The inhibition
 247 of bacterial metabolism is manifested by attenuation of light emittance which corresponds to the toxicity
 248 level of the tested substance (Abbas et al. 2018, Bulich 1982).

249 For the studied mixtures, the curve dose-response for *A. fischeri* at 30 min are shown in Figure 2.
 250 For this biomodel, the values of EC₅₀ are very high varying between 30000-172000 mg/L, indicating a low
 251 toxicity of the studied mixtures towards this biomodel; however, DES containing glucose are more toxic
 252 than DES containing sorbitol, concretely between 4.1 and 2.5 times more toxic, depending on the DES. In
 253 all cases, toxicity decreases with water content. In the case of DES with sorbitol, this decrease of toxicity
 254 is almost linear while for glucose DES, the the decrease is more pronounced if the water content increases.
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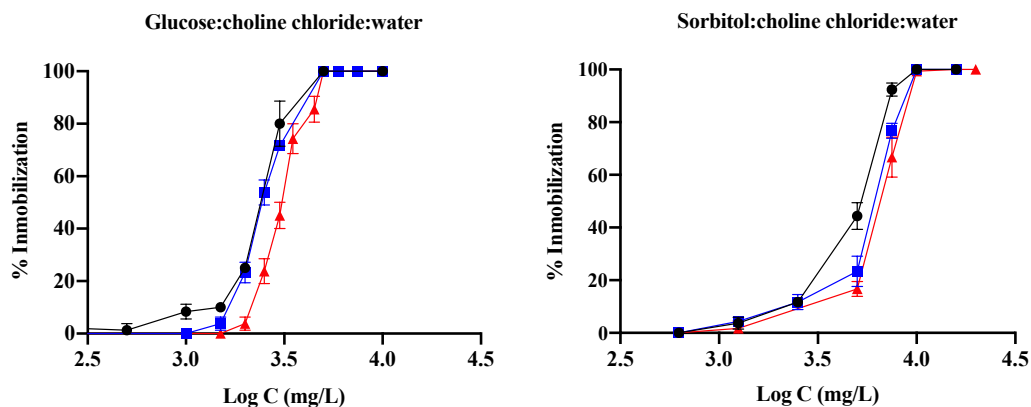
256
 257 Figure 2. Curve dose-response in *A. fischeri* for Glu and Sor at 30 min. Content of: 5 of water (●), 7.5 of
 258 water (■) and 10 of water (▲).

260 *D. magna* is one of the oldest organisms used in biological research (Tkaczyk et al. 2021). They
 261 can filter bacteria, algae, protozoans or other small aqueous particles. *Daphnia* are quite important for the
 262 freshwater aquatic food chain, as a primary consumer for predatory invertebrates and fish. Additionally,
 263 daphnids consume algae and thus, improve water quality (Dietrich et al. 2010, Tkaczyk et al. 2021). The
 264 *Daphnia* motility is easy to observe and therefore, this feature is commonly used in immobilization,
 265 lethality and reproduction tests (Bownik 2020). In addition, swimming behavior parameters (swimming
 266 activity, swimming time, swimming speed, behavioral strength and hopping frequency) can be used as
 267 sensitive endpoints (Bownik 2017). However, it is important to control the assay conditions because

268 variations in water temperature or medium culture can affect to the physiological and physical effects on
269 these organisms(Loiterton et al. 2004).

270 For *D. magna*, the curves dose-response at 24 h for all the studied chemicals are shown in Figure
271 3. Once again, DES containing sorbitol are less toxic than glucose DES (nearly 2 times less toxic). Besides,
272 toxicity decreases in both cases with water content; the behaviour of DES containing sorbitol is linear-like
273 while, for glucose DES the decrease in toxicity is more pronounced.

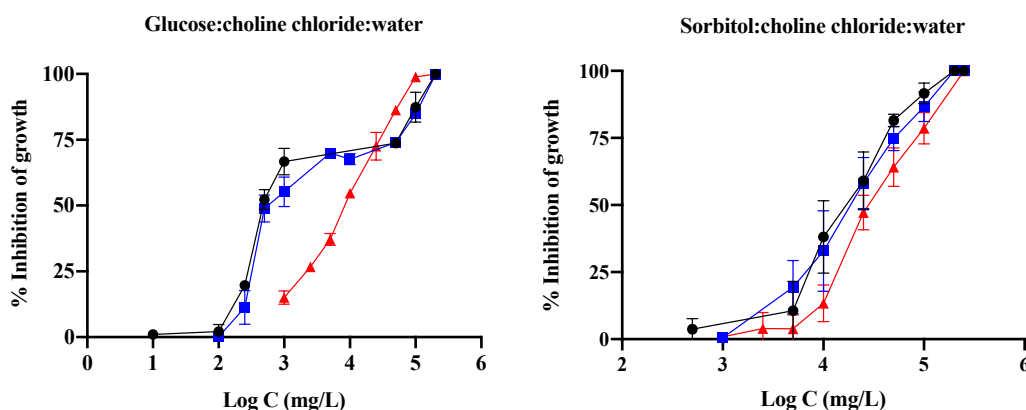
274 Algae play an important role in the maintaining the structure and function of aquatic ecosystems
275 since are primary producers. Algae can be considered as an important bioindicator in the evaluation of the
276 water quality and ecotoxicity of substances(Guo et al. 2020). In particular, *R. subcapitata* is a typical species
277 used in ecotoxicological studies, recommended by the Organization for Economic Cooperation and
278 Development (OECD), because of its global distribution, fast growth, and great sensitivity(Mo et al. 2022).



279
280 Figure 3. Curve dose-response in *D. magna* for Glu and Sor at 24 h. Content of: 5 of water (●), 7.5 of
281 water (■) and 10 of water (▲).

282
283 For *R. subcapitata*, the curves dose-response at 72 h for all studied chemicals are shown in Figure
284 4. As we can observed, the DES formed by glucose and choline chloride show a non-typical sigmoidal
285 curve, especially the case of Glu5 and Glu7.5. Although the effect of a certain DES concentration with
286 exposure time has not been studied, this characteristic profile could be indicative of a diauxic effect in the
287 inhibition growth process (Blaiseau &Holmes 2021). The slope in growth inhibition is very steep until the
288 concentration of DES reaches a value of approximately 1000 mg/L. From there, the inhibition remains
289 constant around 75% in a very wide range of concentrations (from 1,000 to 100,000 mg/L). This behaviour
290 can be related to the viscosity of these mixtures because they are more viscous than Glu10. For this
291 biomodel, toxicity decreases with the water content in the moiety, being the most pronounced decrease in
292 the case of Glu10 DES. DES containing sorbitol are clearly less toxic than glucose DES. These results can
293 be explained in terms of viscosity; Qin *et al.* showed that the motility of algae (*Chlamydomonas reinhardtii*)
294 is influenced by the viscosity of the environment, *i.e.*, the motility of algae is related to a complex interaction
295 between swimmers kinematics and the properties of that material(Qin et al. 2015).

296 Additionally, Coppola *et al.* showed that the velocity and angular diffusion of the algae decreases
297 when the viscosity of the surrounding medium is increased. They also observed that algae do not
298 concentrate in the region of high viscosity despite them swimming slower there(Coppola &Kantsler 2021).
299



300
 301 Figure 4. Curve dose-response in *R. subcapitata* for Glu and Sor at 72 h. Content of: 5 of water (●), 7.5
 302 of water (■) and 10 of water (▲).

304 4. Conclusions

305 In this paper, the ecotoxicity of several DES (Glu5, Glu7.5, Glu10, Sor5, Sor7.5 and Sor10) has
 306 been evaluated in three aquatic biomodels (*A. fischeri*, *D. magna* and *R. subcapitata*). Results indicate a
 307 low ecotoxicity of the studied solvents. For glucose DES, the most sensitive biomodel is *R. subcapitata*,
 308 while for DES containing sorbitol, the most sensitive species is *D. magna*. The highest values of EC₅₀ are
 309 found for the biodel *A. fischeri* for all the studied DES.

310 To explain the results, the kinematic viscosity of the mixtures has been also measured at 25°C.
 311 The toxicity of the DES decreases as the water content increases. This result correlates with the kinematic
 312 viscosity values found for the studied DES. Regarding to the GHS classification, there is no need to classify
 313 for short-term (acute) hazard for all mixtures under study.

315 Funding

316 The PLATON research group acknowledges financial support from Gobierno de Aragón and Fondo Social
 317 Europeo “Construyendo Europa desde Aragón” E31_17R. Furthermore, we thank EEE53 SL and the
 318 business groups Pinares de Venecia División Energética and Brial (ENATICA) for their support. Both
 319 business groups are committed to sustainable developments through environmental respect. Diego
 320 Errazquin and M^a Pilar Garralaga thank Novaltia, Banco Sabadell for her financial support.

322 Compliance with Ethical Standards

323 Competing Interests: the authors have no relevant financial or non-financial interests to disclose.

324 Research involving Human Participants and/or Animals: no human or superior animals have been used in
 325 this work.

326 Informed consent: does not apply in this work.

327 Availability of data and materials: does not apply in this work.

332 **Author Contribution**

333 Conceptualization: Laura Lomba and Beatriz Giner; Formal analysis: Laura Lomba and Beatriz Giner;
334 Investigation: Diego Errazquin, M^a Pilar Garralaga, and Noelia López; Methodology: Laura Lomba; Project
335 administration: Beatriz Giner.; Resources: Laura Lomba.; Supervision: Beatriz Giner.

336

337 **Consent to participate**

338 All authors agreed with the content and that all gave explicit consent to submit.

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341 **Consent to publish**

342 All authors agreed with the publication of the manuscript in Environmental Science and Pollution
343 Research.

344 **References**

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